j9f6cr3rf

Size selection (Purification) (1X ratio) V.(j9f6cr3rf)

Version 1 is forked from Size selection (Purification)

Manage mention

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Abstract

Size selection (Purification)

1 Calculate the amount of magnetic beads you need and **DISTRIBUTE in a new 1.5 mL eppendorf tube (**Mix the bottle **VIGOROUSLY** to make sure it's homogeneous**).**
For example, if you have 5 samples to clean up, distribute 49.5 µl in a new 1.5 mL eppendorf tube.

	A	B
	Sample	**Magnetic beads need**
	1	25 µl
	2	50 µl
	3	75 µl
	4	100 µl
	5	125 µl
	6	150 µl
	7	175 µl
	8	200 µl

Note

The ratio is **25 µl of sample** use **25 µl of magnetic beads** to clean up (1 X to keep moderate to long DNA fragments).

Note

Magnetic bead: BeaverBeads™ DNA Select Isolation

2 In each 25 µL sample, add 25 µL magnetic beads in.

Note

Pipetting thoroughly every time before adding to make sure all magnetic beads mixed well.

3 Mix sample and beads by **gently flicking** the tube for 5mins 00:05:00 5m

Note

Be gentle, mix too aggressive will make DNA fragmented

4 Transfer the tube to the 8-strip magnetic rack.
After all the magnetic beads are arrested, remove the supernatant.

5 Add 200 µL 80% ethanol, flip the magnetic rack around to clean the pellet.
Wait until all the magnetic beads are arrested, remove the supernatant.

> Note
>
> **FRESHLY** make 80% ethanol

6 Repeat the **step 5**.

7 Flash spin the tube, and put on the magnetic rack.
Remove **ANY trace of liquid residuals** using 10 µl pipette.

> Note
>
> Caution: DO NOT let the beads crack

8 Add 15 µL elution buffer.
Mix gently by flicking the tube and make sure all magnetic beads are dissolved in buffer, then flash spin down the sample.

9 Incubate the tube in PCR thermal cycler using "**37_incubation**" program for 10min 00:10:00 . 10m

10 Transfer the tube to the magnetic rack.
After all the magnetic beads are arrested, collect the 15 µL supernatant to a 200 µl PCR tube or a 8-strip PCR tube.

11 The clean-up sample is now ready for 2' PCR.